

## FREE STEROLS OF THE RED ALGA *GIGARTINA SKOTTSBERGII*

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**Key Word Index**—*Gigartina skottsbergii*, Gigartinales, Rhodophyceae, sterols, cholest-5-en-3 $\beta$ -ol

**Abstract**—The free sterols of the red alga *Gigartina skottsbergii* have been identified by means of GC and GC/MS analyses. The mixture contained saturated and unsaturated C<sub>27</sub>, C<sub>28</sub> and C<sub>29</sub> sterols. The major component was cholest-5-en-3 $\beta$ -ol. Cholesta-5,24-dien-3 $\beta$ -ol (desmosterol) was present in low proportion but no side chain hydroxylated components were detected.

### INTRODUCTION

The main sterols of the majority of red algae are C<sub>27</sub> compounds, cholesterol being the major component of most of them, although in a few species desmosterol is present in substantial amounts and may even be the major sterol [1–3]. Red algae also contain, though in minor amounts, C<sub>26</sub>, C<sub>28</sub> and C<sub>29</sub> sterols and in some species the presence of side chain hydroxylated sterols has been

reported [4–6].

We wish to report here the sterol composition of *Gigartina skottsbergii*, a red alga from the south Atlantic ocean.

### RESULTS AND DISCUSSION

Fresh specimens of *G. skottsbergii*, collected at Puerto Madryn, Chubut, were freeze-dried and the residue was

Table 1 Free sterols of the alga *Gigartina skottsbergii*

Sterols	MS characteristic fragments	RR,*	Composition (%)
22- <i>trans</i> -Cholesta-5,22-dien-3 $\beta$ -ol	384 [M] <sup>+</sup> , 366, 351, 300, 273, 271, 255, 213, 111, 69, 55	0.94	0.7
Cholest-5-en-3 $\beta$ -ol	386 [M] <sup>+</sup> , 371, 368, 301, 275, 273, 255, 231, 213, 145, 43	1.00	92.7
5 $\alpha$ -Cholestan-3 $\beta$ -ol	388 [M] <sup>+</sup> , 373, 370, 355, 275, 257, 233, 217, 215, 43	1.02	tr†
22- <i>trans</i> -24 $\xi$ -Methylcholesta-5,22-dien-3 $\beta$ -ol	398 [M] <sup>+</sup> , 380, 365, 300, 271, 255, 213, 69, 55	1.04	1.1
Cholesta-5,24-dien-3 $\beta$ -ol	384 [M] <sup>+</sup> , 369, 366, 351, 300, 299, 282, 271, 253, 231, 229, 213, 211	1.07	2.1
24 $\xi$ -Methylcholest-5-en-3 $\beta$ -ol	400 [M] <sup>+</sup> , 385, 382, 367, 315, 289, 273, 213, 105, 43	1.10	0.8
24 $\xi$ -Methyl-5 $\alpha$ -cholestan-3 $\beta$ -ol	402 [M] <sup>+</sup> , 387, 384, 369, 275, 233, 217, 215, 43	1.12	tr
24-Methylcholesta-5,24(28)-dien-3 $\beta$ -ol	398 [M] <sup>+</sup> , 383, 380, 314, 299, 281, 271, 255, 229, 213, 55	1.14	1.1
22- <i>trans</i> -24 $\xi$ -Ethylcholesta-5,22-dien-3 $\beta$ -ol	412 [M] <sup>+</sup> , 397, 394, 379, 351, 300, 273, 271, 255, 69, 55	1.16	0.5
24 $\xi$ -Ethylcholest-5-en-3 $\beta$ -ol	414 [M] <sup>+</sup> , 396, 381, 368, 329, 303, 255, 213, 91, 55	1.23	0.5
24 $\xi$ -Ethyl-5 $\alpha$ -cholestan-3 $\beta$ -ol	416 [M] <sup>+</sup> , 401, 398, 383, 275, 233, 215, 55	1.25	tr
24-Ethylcholesta-5,24(28)-dien-3 $\beta$ -ol	412 [M] <sup>+</sup> , 397, 394, 314, 299, 281, 273, 271, 255, 229, 213, 55	1.27	0.4
24-Ethyl-5 $\alpha$ -cholest-24(28)-en-3 $\beta$ -ol	414 [M] <sup>+</sup> , 399, 396, 316, 301, 273, 233, 215, 55	1.28	tr

\*RR<sub>i</sub> = retention times of the free sterols relative to cholest-5-en-3 $\beta$ -ol

†tr = traces

extracted with methanol. The residue obtained by evaporation of the solvent was partitioned between ethyl acetate and water. The organic extract was dried and evaporated. The residue was chromatographed on silica gel and the sterol fraction was fractionated further by HPLC. Each subfraction was analysed by GC and GC/MS leading to the identification of the sterols of the mixture (see Table 1) by comparison with known standards. As it has been normally found in red algae, cholesterol was by far the most abundant constituent of the mixture, which also contained  $C_{28}$  and  $C_{29}$  saturated and unsaturated components. It is interesting to note that desmosterol was present, in small amount, in the free state but we were unable to detect any side chain hydroxylated sterol.

#### EXPERIMENTAL

**General.** HPLC was carried out on a Whatman Partisil M9 10/50 ODS-2 column using an RI detector (mobile phase MeOH). GC was on a fused silica capillary column (12 m  $\times$  0.02 mm) coated with methyl silicone fluid (Hewlett-Packard). Computerized GC/MS was on a Varian-Mat CH7-A instrument at 70 eV.

**Plant material.** *Gigartina skottsbergii* was collected at Puerto Madryn, Chubut, Argentina in winter at 8 m depth. Voucher specimens have been deposited at the Centro Nacional Patagónico, Puerto Madryn.

**Extraction and analysis.** Fresh algae (3.4 kg) were freeze-dried and the residue (1.2 kg) was extracted with MeOH (2  $\times$  21) yielding an extract which was partitioned between EtOAc and  $H_2O$ . The syrup obtained by evaporation of the organic solvent (4.45 g)

was chromatographed twice on silica gel columns eluted with toluene and toluene- $CH_2Cl_2$  (1:1) to separate the free sterol fraction. This fraction (99 mg) was purified by HPLC yielding four subfractions that were analysed by GC and GC/MS techniques. Identification of each sterol was made by comparison with the MS data of authentic samples (see Table 1).

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#### REFERENCES

- 1 Goodwin, T. W. (1974) in *Algal Physiology and Biochemistry* (Stewart, W. D., ed.), Botanical Monographs, Vol. 10. Blackwell Scientific Publications, London.
- 2 Goad, L. J. (1978) in *Marine Natural Products: Chemical and Biological Perspectives* (Scheuer, P. J., ed.), Vol. II, Chap. 2. Academic Press, New York.
- 3 Amico, V., Chillemi, R., Sciuto, S., Tringali, C., Cormaci, M. and Furnari, G. (1982) *Naturalista Sicil.* S. IV, VI (Suppl.), 1, pp. 95–106.
- 4 Combaut, G., Codomier, L., Teste, J. and Pedersen, M. (1981) *Phytochemistry* 20, 1748.
- 5 Francisco, C., Combaut, G., Teste, J., Tarchini, C. and Djerassi, C. (1979) *Steroids* 34, 163.
- 6 Kabore, S. A., Combaut, G., Vidal, J. P., Codomier, L., Passet, J., Girard, J. P. and Rossi, J. C. (1983) *Phytochemistry* 22, 1239.

## ANTHRAQUINONES FROM *CASSIA SOPHERA* ROOT BARK

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**Key Word Index.**—*Cassia sophera*, Leguminosae, 1,8-dihydroxy-3,6-dimethoxy-2-methyl-7-vinyanthraquinone, 1,3-dihydroxy-5,7,8-trimethoxy-2-methylantraquinone

**Abstract.**—Two new anthraquinones have been isolated from the root bark of *Cassia sophera* and characterized as 1,8-dihydroxy-3,6-dimethoxy-2-methyl-7-vinyanthraquinone and 1,3-dihydroxy-5,7,8-trimethoxy-2-methylantraquinone.

*Cassia sophera* (Leguminosae) is well known for its medicinal value [1], roots, flowers and heartwood have been studied chemically [2–6].

From the benzene fraction of the acetone extract of the root bark of *Cassia sophera* two new anthraquinones have now been isolated and characterized by spectral and

chemical studies. Both compounds (1 and 2) responded to characteristic colour tests for anthraquinones. Their  $\lambda_{\max}$  and IR spectra supported the above conclusion. Strong peaks at  $1460\text{ cm}^{-1}$  in the IR spectrum of 1 and at  $1450\text{ cm}^{-1}$  in that of 2, a signal at  $\delta 2.35$  in the  $^1\text{H}$  NMR spectrum of 1 and 2.38 in that of 2 and identification of 2-